Low-dose radiation: Thresholds, bystander effects, and adaptive responses

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Ionizing radiation fills the universe. Daily ionizing particles and rays collide with molecules in ~1% of the 100 trillion cells that make up the average human. These collisions generate clusters of free radicals known as reactive oxygen species that randomly damage cellular constituents including DNA (1). Certain types of ionizing radiation are more effective at generating reactive oxygen species; one α-particle is at least 10 times more damaging than one γ-ray. To take these differences into account, the Sievert (Sv), a unit that multiplies the absorbed dose in grays (Gy) by the relative effectiveness of the particle or ray to inflict damage, was developed. On this scale, natural background radiation is ~0.01 mSv/day, although there are areas on earth that have values 5-fold higher (2), and space-station inhabitants may receive ~1 mSv/day (3). At the other end of the scale, acute exposures of >150 mSv, a range known as high-dose radiation, have measurable and often serious immediate effects on humans (4). Between background and high-dose radiation is the range of exposures known as low-dose radiation. Low-dose radiation has no immediately noticeable effects on humans; nevertheless there is great interest in its long-term biological effects, which may include cancer in exposed individuals and genetic defects in their progeny.

Research into the biological effects of low-dose radiation exposure is hindered by a lack of assays sensitive enough to measure the relevant cellular alterations. More-sensitive assays are being developed, an important one being the ability to detect the cellular presence of the most serious and potentially lethal type of cellular damage, the DNA double-strand break (DSB). This assay is based on the finding that one of the highly conserved histone proteins that package the DNA into chromatin, H2AX, becomes phosphorylated at the sites of nascent DNA DSBs (5–8). The response is highly amplified and rapid, involving the phosphorylation of hundreds to thousands of H2AX molecules within minutes on several megabase equivalents of chromatin flanking the DSB. When visualized with an antibody, the phosphorylated H2AX species, named γ-H2AX, appears as nuclear foci (Fig. 1). In this issue of PNAS, Rothkamm and Löbrich (9) report that the number of DSBs formed, as measured by the number of γ-H2AX foci formed, is linear with dose from 1 mGy to 2 Gy and is also in line with pulse-field gel electrophoresis measurements at higher doses. Greatly extending previous quantitative measurements (10), their findings demonstrate that γ-H2AX focus formation is several orders of magnitude more sensitive than other current methods for detecting DSBs (7).

How γ-H2AX foci function in DNA DSB repair and rejoining is poorly understood, but the foci do serve to recruit DNA-repair proteins to DSB sites (11). γ-H2AX foci also form as part of normal cellular processes involving DSBs, including homologous recombination during meiosis and genetic recombination during immune system development (12–15); mice lacking H2AX are viable but deficient in these two areas as well as being sensitive to ionizing radiation. Elucidating the biological roles of γ-H2AX foci will bring greater understanding into the various cellular mechanisms for DNA DSB formation and repair.

Rothkamm and Löbrich (9) used amounts of γ radiation as low as 1 mGy, a dose that generates on average one track of clustered reactive oxygen species per nucleus and thus is considered the lowest dose that can affect a whole cell culture or animal. With 1 mGy, ~3% of irradiated cells sustain a DNA DSB. Compared with metabolic DSBs, radiation-induced DSBs are more hetero-
certain doses of low-dose radiation may even be beneficial. Typically the adaptive response is induced with 1–100 mGy of γ-rays, doses 100–10,000 times larger than the natural background of ~0.01 mSv/day. This model was first proposed in 1984 to explain the finding that cultures of human lymphocytes growing in low concentrations of radioactive thymidine developed fewer chromosomal aberrations than cultures of nonradioactive lymphocytes when both were challenged with high-dose radiation (22). Other studies also seem to support this model (23–26, ‡).

Yet another model is the bystander-effect model, which postulates that low-dose radiation may be even more damaging than that predicted by the linear nonthreshold model. For example, in cell cultures irradiated so that only 1% of the cells sustained a collision with an α-particle, sister chromatid exchanges were observed in >30% of the cells (27). Other studies have also supported this model (28–31). These effects suggest that irradiated cells may signal their distress to other cells, perhaps by direct cell-to-cell interaction or by molecules secreted into the medium. The latter form of cellular communication is supported by findings showing that the bystander effect could be induced in nonirradiated cell cultures incubated with conditioned medium from irradiated cultures (32, 33). One possible candidate for the signal is IL-8 (34), a cytokine with important roles as a chemoattractant and activator of polymorphonuclear leukocytes that is up-regulated and secreted in a variety of cell types during oxidative stress.

The adaptive response and the bystander effect can occur in the same experimental system. When cultures of C3H10T½/ cells were irradiated through a microbeam with known numbers of α-particles, survival was less (65%) than that predicted from the linear nonthreshold model (90%), which is a bystander effect. However, when the cultures were irradiated with 20 mGy of γ-rays 6 h before α-particle exposure, survival was increased (75%), which is an adaptive response (35). This study points to what may be important differences in the two phenomena. The bystander effect is typically induced by the more-damaging α-particles, whereas the adaptive response is typically induced with γ-rays. 


Quiescent cells signal their distress to their neighbors through secretion of IL-8 and other signaling molecules.

Rothkamm and Löbrich (9) report findings that also support a cellular communication model. In studies examining the incidence of DSBs in quiescent cultures of normal human fibroblasts after low-dose radiation, the expected numbers of DSBs, proportional to dose, were found at 3 min. Over 24 h the numbers decreased; however, they did not return to the preexposure average of 0.05 DSB per cell but stabilized at 0.1 DSB per cell for at least 14 days independent of dose. Thus 5% of the cells in the unirradiated cultures contained DSBs compared with 10% in the exposed cultures. However, when the quiescent cultures were exposed daily to 1.2 or 5 mGy for 10 days, the percentage of cells sustaining DSBs returned in each case to 10% within 24 h of the last irradiation. Thus only the first exposure increased the percentage of cells with DSBs after 24 h; subsequent exposures had no effect. When irradiated quiescent cultures were subcultured to induce growth, the percentage of cells with DSBs returned to the basal level of 5% at 7 days. However, apoptotic cells and cells with micronuclei were more abundant in the growing cultures derived from irradiated quiescent cultures than in those derived from the controls. In addition, cell survival after 200 and 1.2 mGy (80% and 90%, respectively) was significantly less than that predicted from the linear nonthreshold model (93% and 99+%).

To explain their findings, the authors suggest that quiescent cells signal their distress to their neighbors, perhaps as previously discussed, through secretion of IL-8 and other signaling molecules. Concerning the increased apoptosis and decreased survival in cell populations derived from the irradiated cultures, the authors suggest that these responses may be beneficial by eliminating injured or less-healthy cells from organisms subjected to low-dose irradiation. If cultures of quiescent normal human fibroblasts are a valid model system, then similar biological effects may occur in...
organs of animals subjected to low-dose radiation.

Understanding the biological consequences of exposures to low-dose radiation is becoming increasingly important for humans and other organisms as greater exposures to ionizing radiation occur from new man-made sources and space travel. If doses 100 times back-ground are harmless (threshold model) or even beneficial (adaptive-response model), exposure standards could be relaxed, resulting in substantial savings. In addition, the adaptive response and bystander effect could prove useful in cancer therapy if normal and tumor cells are found to respond differently. However, until the mechanisms of these two phenomena are understood and their effects become predictable, few would propose a relaxation of standards. This contribution of Rothkamm and Löbrich (9) is a significant advance toward the elucidation of the immediate as well as long-term biological consequences of low-dose radiation and their effects on human health and safety.